

Cas9 Nuclease

Catalog # PR-137210

Description	Recombinant enzyme purified from <i>E.coli</i> . Cas9 Nuclease is an RNA-guided endonuclease that catalyzes site-specific cleavage of double stranded DNA.
Source	An <i>E. coli</i> strain that carries recombinant Cas9 gene from <i>Streptococcus pyogenes</i>
Size	50 µg/ 50 µl
Physical Appearance	Colorless solution.
Formulation	100µg in 100µl of 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM DTT, 300 mM NaCl, and 50% (v/v) Glycerol.
Stability	All components are stable for 1 year from the date of shipping when stored and handled properly. Store all components at -20°C. Avoid repeated freeze-thaw cycles of all components to retain maximum performance.
Purity	Greater than 95.0% as determined by SDS-PAGE analysis.
Reaction Buffer (10x)	200 mM HEPES, 50 mM MgCl ₂ , 1 M NaCl, 1 mM EDTA, pH 6.5
Usage	Novateinbio's products are furnished for LABORATORY RESEARCH USE ONLY. The product may not be used as drugs, agricultural or pesticidal products, food additives or household chemicals.

Protocol

The protocol listed below is for reference only. The user may optimize the protocol according to their own experiments.

In vitro Cas9 mediated digestion of DNA

1. Add the following components to a sterile, nuclease-free tube on ice:

Components	Volume	Final Concentration
sgRNA (300 nM)	2 μ l	~30 nM
Cas9 Nuclease Protein (1000 nM)	0.60 μ l	~30 nM
10X Cas9 Nuclease Reaction Buffer	2 μ l	1X
Nuclease-free H ₂ O	12.4 μ l	-
Pre-Incubate for 15 minutes at room temperature		
Substrate DNA (30 nM)	3 μ l	3 nM
Total Volume	20 μ l	-

2. Collect all components by a brief centrifugation. Incubate the reaction at 37 °C for 1 hour.
3. Analyze fragments by running an agarose gel electrophoresis.

RNP Complex Formation

Gently mix the reaction and incubate at room temperature for 10 minutes. Then place on ice for following transfection by electroporation or liposome.

Components	Volume	Final Concentration
sgRNA (1000 nM)	1.2 μl	~120 nM
Cas9 Nuclease Protein (1000 nM)	1.2 μl	~120 nM
Opti-MEM	12.6 μl	-
Total	15 μl	

Trypsinize and Prepare HEK293T Cells

1. Seed the cells so that they will be around 70-90% confluent on the day of transfection.
2. During the RNP/liposome incubation, trypsinize the cells, washing once to remove any traces of trypsin. Resuspend the cells in 10 ml of media and count.
3. Calculate the dilution and volume needed to get the cells to 3.2×10^5 cells per ml. You will need 125 μl of cells per well.

Transfect Cells with Liposome Complexes

1. From each tube of RNP/liposome complex, aliquot 25 μl into 3 wells of a 96-well plate.
2. Add 125 μl of cells (3.2×10^5 cells/ml) to each well containing RNP/liposome complex and pipette up and down gently a few times.
3. Incubate the cells in a humidified 37°C, 5% CO₂ incubator for 48-72 hours.

Harvest DNA and Amplify Target Region

1. Gently aspirate the media from the cells and wash twice with 100 μl 1X PBS.
2. Add 75 μl of DNA Extraction Solution and shake/vortex for 5 minutes. Transfer the solution to a PCR plate or tubes and place in a thermocycler, running the following program:

65°C for 15 min
95°C for 15 min
Hold at 4°C

3. Dilute the DNA 1:10 in nuclease-free water.
4. Follow the protocol detailed in the T7E1 assay kit

Quality Control:

Exonuclease, endonuclease and non-specific nuclease activities not detected.